AD-A238 544

AD_____

WORLD REFERENCE CENTER FOR ARBOVIRUSES AND RETROVIRUSES

DTIC FCTE JUL 19 1991 FINAL REPORT

ROBERT E. SHOPE

MAY 1, 1990

20030211156

Supported by

U.S. ARMY MEDICAL RESEARCH AND DEVELOPMENT COMMAND Fort Detrick, Frederick, Maryland 21701-5012

Grant No. DAMD17-87-G-7005

Yale University School of Medicine New Haven, Connecticut 06510

Approved for public release; distribution unlimited.

The findings in this report are not to be construed as an official Department of the Army position unless so designated by other authorized documents

91-05589

Š	Ē	C	U	jŘ	Ī	īŸ	7	LA	33	ΊF	īζ	A	1OI	V	ŌĒ	THIS	PA	GΕ

		RE	PORT I	DOCUMENTATIO	N PAGE			Form Approved OMB No. 0704-0188	
1a. REPORT SI Uncla	ECURITY CLAS	SSIFICATION	4		16. RESTRICTIVE MARKINGS				
2a. SECURITY	CLASSIFICATION	ON AUTHO	RITY		3. DISTRIBUTION				
20. DECLASSIF	ICATION / DO	WNGRADIN	IG SCHEDU	LE	Approved for public release; distribution is unlimited.				
4. PERFORMIN	G ORGANIZA	TION REPO	RT NUMBE	R(S)	5. MONITORING ORGANIZATION REPORT NUMBER(S)				
6a. NAME OF Yale Univ				6b. OFFICE SYMBOL (If applicable)	7a. NAME OF M	ONITORING ORGA	NIZATION		
6c. ADDRESS (7b. ADDRESS (City, State, and ZIP Code)				
New Haven	, Connect	ticut 06	6510						
Sa. NAME OF			اممانه	Bb. OFFICE SYMBOL	9. PROCUREMEN	I INSTRUMENT ID	ENTIFICATION	ON NUMBER	
Research	tion U.S. & Develop			(If applicable)	DAMD17-87-G-7005				
Sc. ADDRESS (C	ity, State, and	d ZIP Code)			10. SOURCE OF FUNDING NUMBERS				
Fort Detr	ick. Fred	ierick.	MD 2170	01-5012	PROGRAM ELEMENT NO.	PROJECT NO. 3M1-	TASK NO.	WORK UNIT ACCESSION NO.	
	,				61102A	61102BS13	AA	DA312093	
12. PERSONAL Robert E. 130. TYPE OF R Final	AUTHOR(S) Shope, M	1. D .	o. TIME CO	viruses and Ret	14 DATE OF REPOR 1990 May 1	RT (Year, Month, I	Day) 15.	PAGE COUNT	
17.	GROUP	CODES SUB-GR	IOUP	18. SUBJECT TERMS (C Arbovirus, ret					
26	13			ELISA, yellow	•	i sarcoma,	rapid d	iagnosis,	
06	03			flavivirus, R					
The World Reference Center for Arboviruses and Retroviruses received viruses from the United States and foreign countries for characterization and identification. New viruses, or viruses causing diseases not previously recognized included a subtype of Cache Valley virus from a febrile military recruit, LaCrosse virus from encephalitic dogs, Kagoshima and Cache Valley viruses associated with fetal abnormalities of livestock, and new phleboviruses from tropical America and West Africa. Genetically engineered flavivirus trpE proteins reacted specifically by ELISA. The cell lysate antigen technique for ELISA was adapted to a large number of arboviruses. ELISA was also developed for rapid testing of 17D yellow fever vaccinees. Limited primer extension sequencing of dengue and Japanese encephalitis viruses showed that their genotypes were geographically focal. The method was used to demonstrate the origin of an introduced dengue virus. Retrovirus isolation capability was developed and a novel enveloped agent from tissues of Kaposi sarcoma patients was studied serologically. Reference reagents were distributed to scientists in over 20 countries. **O DISTRIBUTION/AVAILABILITY OF ABSTRACT** **DUNCLASSIFIEOTUNIUMITED** **DOINCLASSIFIEOTUNIUMITED** **TOTIC USERS** **PORT OF RESPONSIBLE INDIVIDUAL** **TOTIC USERS** **TOTIC USERS** **PORT OF RESPONSIBLE INDIVIDUAL** **TOTIC USERS** **TOTIC									
220. NAME OF R			Ĺ		226 TELEPHONE (M	clude Area Code)			
O form 1473,	inces Bos	cian		Previous editions are o	301/663-732 bsolete			-RMI-S ION OF THIS PAGE	

ABSTRACT

The World Reference Center for Arboviruses and Retroviruses received viruses from the United States and foreign countries for characterization and identification. New viruses, or viruses causing diseases not previously recognized included a subtype of Cache Valley virus from a febrile military recruit, LaCrosse virus from encephalitic dogs, Kagoshima and Cache Valley viruses associated with fetal abnormalities of livestock, and new phleboviruses from tropical America and West Africa. Genetically engineered flavivirus trpE proteins reacted specifically by ELISA. The cell lysate antigen technique for ELISA was adapted to a large number of arboviruses. ELISA was also developed for rapid testing of 17D yellow fever vaccinees. Limited primer extension sequencing of dengue and Japanese encephalitis viruses showed that their genotypes were geographically focal. The method was used to demonstrate the origin of an introduced dengue virus. Retrovirus isolation capability was developed and a novel enveloped agent from tissues of Kaposi sarcoma patients was studied serologically. Reference reagents were distributed to scientists in over 20 countries.

Accesio	ni For							
NTIS	CEASI	V						
Disc	-	١.						
10 10	-	(.)						
Ju 11.11.	Justination							
Ву								
Di-Libation/								
	-							
Availability Chites								
Dist	Aval. a							
0.31	် ပူင	U.						
Λ								
4-1								

FOREWORD

In conducting the research described in this report, the investigators adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and Use of Laboratory Animals of the Institute of Laboratory Animal Resources, National Research Council (DHEW Publication No. (NIH) 86-23, Revised 1985).

For the protection of human subjects the investigators have adhered to policies of applicable Federal Law 45CFR46.

BODY OF REPORT:

Introduction. The World Reference Center for Arboviruses was established at the Yale Arbovirus Research Unit in 1965 as an outgrowth of The Rockefeller Foundation program on arboviruses which was moved in 1965 to Yale University from New York City. The U.S. Army has supported this program since 1972, initially through joint Navy-Army funding, then through separate contracts and grants, and during the past three years by this grant. The progress of the past three years is included in this report; it covers the work for the entire project which received support from WHO and NIH in addition to that of this grant.

<u>Virus identification</u>. Viruses were identified from the United States and from Panama, Angola, Czechoslovakia, Indonesia, Ivory Coast, Viet Nam, Thailand, Taiwan, Japan, and Brazil. Among these were:

BUNYAVIRUSES. A virus isolated from blood of a febrile U.S. Army recruit on jungle training in Panama was identified as a subtype of Cache Valley virus.

PHLEBOVIRUSES. Eight new phleboviruses were characterized serologically. Odenisrou, a new phlebovirus from the Ivory Coast, was also characterized serologically.

ORBIVIRUSES. A Palyam group virus, Kagoshima, was identified from <u>Culicoides</u> midges from Japan for the first time. This virus was associated with congenital abnormalities in livestock. The virus was shown to be the same as Kasba virus from India.

POXVIRUSES. Ectromelia virus was found in viruses from Czechoslovakia submitted for identification.

Classification of arboviruses. Studies of two serogroups in the genus Orbivirus led to a proposal for defining species. The correlation between the degree of RNA-RNA hybridization and genetic reassortment of these double stranded RNA viruses was excellent.

<u>Diagnosis</u> of disease. Cache Valley virus was associated with an epizootic in sheep of arthrogryposis and hydranencephaly affecting nearly 20% of offspring in Texas and Nebraska. This is the first time a bunyavirus has been implicated in this disease in the Americas.

A virus was isolated by scientists of the University of Georgia from two puppies in rural Georgia. The puppies died of encephalitis. Neutralization tests confirmed the identity of the virus as LaCrosse virus, a cause of human encephalitis. This is the first time this virus has been isolated in nature from encephalitic dogs.

An outbreak of hemorrhagic fever in Pakistan was studied. Although seroreactivity in one patient to Crimean-Congo hemorrhagic fever virus was detected, this virus was apparently not the only cause of the illness.

Characterization of monoclonal antibodies. Monclonal antibodies were developed and characterized for vesicular stomatitis (VSV), Indiana, Cocal, and Alagoas viruses. These were field tested for their diagnostic specificity at the Foot and Mouth Disease Laboratory in Rio de Janeiro.

Semliki Forest virus monoclonal antibodies were used to find a conserved epitopic region on the alphavirus nucleocapsid protein. A mixture of monoclonal antibodies to Semliki Forest virus was blended to develop a sensitive antigen capture ELISA for alphaviruses.

Development of new techniques. Flavivirus trpE fusion proteins were used as highly specific diagnostic reagents in ELISA. A region of the flavivirus RNA, coding for an antigenic domain that is relatively serotype specific, was located between amino acids 300 and 400 of the E protein. Fusion proteins corresponding to this region were produced for Japanese encephalitis and dengue-1 using previously cloned cDNA. Analagous protein for dengue-2 virus was made using the polymerase chain reaction technology and primers derived from published sequences. The fusion proteins were coated directly on the solid phase for use as ELISA antigens. These proteins were not very immunogenic to mice, but were useful in distinguishing among flaviviruses using antibodies raised in mice.

The yellow fever ELISA was adapted to test 17D vaccinees who were immunological virgins for flaviviruses. The ELISA was as sensitive as the plaque reduction neutralization test (PRNT). ELISA showed seroconversion earlier than PRNT in many cases.

Human-pathogenic arboviruses were adapted to a rabbit kidney continuous cell line that used medium containing rabbit sera, in order to develop a system of immunizing rabbits free of heterologous species reactivity. The immune rabbit sera were used in ELISA as coating antibody to capture antigen.

A rapid method of raising murine antibodies utilized intrasplenic inoculation of mice. Antibody was detected by ELISA as early as 3 days post inoculation, and consistently by the fifth day post inoculation.

The utility of the cell lysate method of producing antigens was confirmed. This technique was originated at USAMRIID. Comparison of results of Rift Valley fever cell lysate antigen with conventional mouse liver antigen in tests of human sera from Ethiopia showed a high degree of correlation.

Molecular epidemiology. Limited primer extension sequencing of dengue-1, dengue-2, and Japanese encephalitis viruses has shown geographic clustering of each of these flaviviruses. Forty-six isolates of Japanese encephalitis virus from various sources and localities in Asia were examined for genetic diversity. The isolates segregated into three geographic patterns. One pattern encompassed southern Thailand, Malaysia, and Indonesia and correlated with the zone that has endemic transmission without major Japanese encephalitis epidemics. It is now possible to trace the origin of new cases and of outbreaks of diseases caused by the three viruses.

A strain of dengue-l isolated from the blood of a patient in Angola was genotyped by this technique and found presumptively to be of Caribbean, not African, in origin.

Collection of low passage arbovirus strains. A large collection of low passage arbovirus strains has been developed and maintained lyophilized. Priority was given to yellow fever, dengue, chikungunya, California encephalitis, Venezuelan encephalitis, St. Louis encephalitis, western encephalitis, eastern encephalitis, Japanese encephalitis, and other human disease arboviruses. The original (or as close to original as was available) material was passaged once in C6/36 mosquito cells or in Vero cells. The resulting stock was lyophilized in aliquots. These were stored and distributed to any and all persons requesting material for study. The collection now contains in excess of 400 strains.

Flavivirus sequence data bank. The sequences of flaviviruses are now on line in an electronic data bank and can be accessed by phone. Floppy discs containing the data were mailed to several investigators working with flaviviruses.

Studies of retroviruses. Attempts to identify by IFA a novel enveloped agent isolated from tissues of patients with Kaposi's sarcoma were negative with a large battery of arboviruses. Retrovirus (HIV) isolation capability was established using H-9 and EBV-transformed lymphocyte cell lines. Some of these lines were supplied to the Instituto Nacional de Salud in Bogota to transfer the technology.

<u>Distribution of reagents</u>. Reference reagents including virus stocks, antibodies, antigens, cell lines, and live insects were distributed in the United States and to scientists in more than 20 foreign nations. Data on reagents available for distribution were entered in a dBase-3+ data bank. More than 4,000 entries have been made so far.

PUBLICATIONS

- Arroyo, J.I., Apperson, S.A., Cropp, C.P., Marafino, B.J., Monath, T.P., Tesn, R.B., Shope, R.E. and Garcia-Blanco, M.A. Effect of human gamma interferon on yellow fever virus infection. Am. J. Trop. Med. & Hyg. 38:647-650, 1988.
- Beaty, B.J., Calisher, C.H. and Shope, R.E. Arboviruses, in Diagnostic Procedures for Viral Rickettsial, and Chlamydial Infections, 6th edition, N.J. Schmidt and R.W. Emmons (eds.). Am. Pub. Health Assn., Washington, DC, pp. 797-855, 1989.
- Bilsel, P.A., Tesh, R.B. and Nichol, S.T. RNA genome stability of Toscana virus during serial transorarial transmission in the sandfly <u>Phlebotomus</u> perniciosus. Virus Research 11:87-94, 1988.
- Bodkin, D.K. and Knudson, D.L. Genetic relatedness of Colorado tick fever virus isolates by RNA-RNA blot hybridization. J. Gen. Virol. 68:1199-1204, 1987.
- Brown, S.E., Gonzalez, H.A., Bodkin, D.K., Tesh, R.B. and Knudson, D.L. Intra- and inter-serogroup genetic relatedness of orbiviruses. II. Blot hybridization and reassortment in vitro of epizootic haemorrhagic disease serogroup, bluetongue type 10 and \overline{Pata} viruses. J. Gen. Virol. 69:135-147, 1988.
- Brown, S.E., Morrison, H.A., Buckley, S.M., Shope, R.E., and Knudson, D.L. Genetic relatedness of the Kemerovo serogroup viruses: I. RNA-RNA blot hybridization and gene reassortment in vitro of the Kemerovo serocomplex. Acta Virologica, 32:369-378, 1988.
- Brown, S.E. and Knudson, D.L. Characterization and identification of arthropod cell lines. In "Arboviruses in Arthropod Cells in Vitro," C. Yunker, ed., CRC Press, Boca Raton, pp. 53-56, 1987.
- Brown, S.E., Miller, B.R., McLean, R.G., and Knudson, D.L. Co-circulation of multiple Colorado tick fever virus genotypes. Am. J. Trop. Med. Hyg. 40:94-101, 1989.
- Brown, S.E., Morrison, H.G., Buckley, S.M., Shope, R.E., and Knudson, D.L. Genetic relatedness of the Kemerovo serogroup viruses: I. RNA-RNA blot hybridization and gene reassortment in vitro of the Kemerovo serocomplex. Acta Virol. 32:369-378, 1988.
- Brown, S.E., Morrison, H.G., and Knudson, D.L. Genetic relatedness of the Kemerovo serogroup viruses: II. RNA-RNA blot hybridization and gene reassortment in vitro of the Great Island serocomplex. Acta Virol. 33:206-220, 1989.
- Calisher, C.H., Karabatsos, N., Dalrymple, J.M., Shope, R.E., Porterfield, J.S., Westaway, E.G. and Brandt, W.E. Antigenic relationships between flaviviruses determined by cross-neutralization tests with polyclonal antisera. J. Gen. Virol. 70:37-43, 1989.

Calisher, C.H., Karabatsos, N., Zeller, H., Digoutte, J.P., Tesh, R.B., Shope, R.E., Travassos da Rosa, A.P.A. and St. George, T.D. Antigenic relationships among rhabdoviruses from vertebrates and hematophagous arthropods. Intervirology 30:241-257, 1989.

Calisher, C.H., Monath, T.P., Sabattini, M.S., Mitchell, C.J., Lazuick, J.S., Tesh, R.B., and Cropp, C.B. Isolation of a newly recognized vesiculovirus, Calchaqui virus, and subtypes of Melao and Maguari viruses from Argentina, with serologic evidence for infections of humans and horses. Am. J. Trop. Med. Hyg. 36:114-119, 1987.

Calisher, C.H. and Shope, R.E. Bunyaviridae, in The Laboratory Diagnosis of Infectious Diseases: Principles and Practices, Chap. 34, Springer-Verlag, New York, 1989.

Calisher, C.H., Shope, R.E. and Walton, T.E. Cell cultures for diagnosis of arbovirus infections in livestock and wildlife. J. Tissue Cult. Meth. 11:157-163, 1988.

Clark, G.G., Calisher, C.H., Crabbs, C.L., Canestorp, K.M., Tesh, R.B., Bowen, R.A., and Taylor, D.E. Malpais Spring virus: a new vesiculovirus from mosquitoes collected in New Mexico and evidence of infected indigenous and exotic ungulates. Am. J. Trop. Med. Hyg. 39:586-592, 1989.

Downs, W.G. and Shope, R.E. Yellow fever, In: Handbook of Viral and Rickettsial Hemorrhagic Fevers, J.H.S. Gear, editor, pp. 73-83, CRC Press, Boca Raton, Florida, 1988.

Dutary, B.E., Petersen, J.L., Peralta, P.H. and Tesh, R.B. Transovarial transmission of Gamboa virus in a tropical mosquito, <u>Aedeomyia squamipennis</u>. Am. J. Trop. Med. Hyg. 40:108-113, 1989.

Figueiredo, L.T.M. and Shope, R.E. An enzyme immunoassay for dengue antibody using infected cultured mosquito cells as antigen. J. Virol. Methods 17:191-198, 1987.

Gibbs, E.P.J., Calisher, C.H., Tesh, R.B., Lazuick, J.S., Bowen, R. and Greiner, E.C. Bivens Arm virus: a new rhabdovirus isolated from Culicoides insignis in Florida and related to Tibrogargan virus from Australia. Vet. Microbiol. 19:141-150, 1989.

Gonzalez, H.A. and Knudson, D.L. Genetic relatedness of Corriparta serogroup viruses. J. Gen. Virol. 68:661-672, 1987.

Gonzalez, H.A. and Kundson, D.L. <u>Orbivirus</u> species and speciation: Genetic reassortment between Corriparta serogroup viruses. Intervirology 28:126-133, 1987.

Gonzalez, H.A. and Knudson, D.L. Intra- and inter-serogroup genetic relatedness of orbiviruses: I. Blot hybridization of viruses of the Australian serogroups. J. Gen. Virol. 69:125-134, 1988.

Greiser-Wilke, I., Moenning, V., Kaaden, O.R., and Figueiredo, T.M. Most alphaviruses share a conserved epitopic region on their nucleocapsid protein. J. Gen. Virol. 70:743-748, 1989.

Kew. O.M., Nottav. B.K., Rico-Hesse, R., and Pallansch, M.A. Molecular epidemiology of wild policytrus transmission. Appl. Virol. Res. 2:231-240, 1989.

Knudson, D.L. Nucleic soid hybridization and zoogeography of some Australian orbiviruses. In "4th Australian Arbovirus Symposium, Brisbane, Queensland, Australia, 9 May 1986," T.D. St.George, ed., 1987.

Kreutzer, R.D., Moct. 7.8., Tesh, R.B. and Young, D.B. Brain cell-karyotypes of six species of New and Old World sand flies (Diptera: Psychodidae). J. Med. Ent. 24:609-612, 1987.

Mason, P.W. Maturattin of Japanese encephalitis virus glycoproteins produced by infector mammalian and mesquito cells. Virology 169:354-364, 1989

Mason, P.W., Dairespie, J.M., Gentry, M.C., McCown, J.M., Hoke, C.H., Purke, D.S., Fournier, M.I. and Mason, T.L. Molecular characterization of a neutralizing ismain of the Japanese encephalitis virus structural givenprotein. J. Gen. Virol. 70:207-2049, 1969.

Mergan, J.M., Yedingtechnig, R.J., Peleg, R.A., Shy, J., Peters, C.J., Walker, J.S. and Shope, R.F., Engine-linked impunosorbent assay for detection of artibodies to Pift Valley fever virus in ovine and bovine sers. As. J. Vet. Pessarch, 48:1138-1141, 1987.

Oprands, 7.3. Improved engame-linked immunosorbent assay for the detection of orbivirus antigens by treatment with sodium dodecyl sulfate. Diagn. Microbiol. Infect. Diagn.

Optendy, 1.1. Schwan, J.G., and Main, A.J. Turk-borne Eemerovo group orbiviruses in a Newfoundland seabird colony. Can. J. Microbiol., 34:782-

(b) this, F., Miseri, N., Peralta, P.H. and Tesh, R.B. A human hasenos en ophalistic assertated with residuar stimatitis visus (Indiana serotype) infection. So. 1. Tesp. Med. & Pvg. 20:312-214, 1968.

Piteirn, T.M. I., Midi, J.B., and Teah, P.B. Calivary appraise activity of some fit enric phietotherns hand fites. Insect Bioches, 19:409-412, 1989.

Poblish. N.M. L. Nachereau, A., Modi, D.M., and Teah, P.B. A cover perform from the salisant glants of the sand Claimaria long.palpis is a potent was collater. The constant public is a

on ja. P.P. ann Jaso, P.P. Mara of our fortished wingsam which infant Nationalists. The Mora Phanes of Same P. Pracesal Windows and P.P. Magner. aditors. Flanca, New York, 1987, pp. 1884-194.

Note, P.F., words'', C.E. and Tracase and Press, A.P.P.A. The epidemi logs of discourses in sedite express in an aps Card Guama. The crasticidae . The Fy feat of grand Arthropolic results in Discusses. T.P. Marsh, Phina. C. F. Cress, Bura Parin, Planta, 1986.

- Shope, R.E. African hemorrhagic fever. In "Cecil Textbook of Medicine," J.B. Wyngaarden and L.H. Smith, Jr., eds., Chap. 366, 1987.
- Shope, R.E. Hemorrhagic diseases caused by arenaviruses. In "Cacil Textbook of Medic J.B. Wyngaarden and L.H. Smith, Jr., eds., Chap. 365, 1987.
- Shope, R.E. Crimean-Congo hemorrhagic fever. In "Cecil Textbook of Medicine," J.B. Wyngaarden and L.H. Smith, Jr., eds., Chap. 364, 1987.
- Shope, R.E. Tick-borne flavivirus diseases: Kyasanur Forest disease and Omsk hemorrhagic fever. In "Cecil Textbook of Medicine," J.B. Wyngaarden and L.H. Smith, Jr., eds., Chap. 363, 1987.
- Shope, R.E. Hemorrhagic fever caused by dengue viruses. In "Cecil Textbook of Medicine," J.B. Wyngaarden and L.H. Smith, Jr., eds., Chap. 362, 1987.
- Shope, R.E. Arthropod-borne viral diseases. Introduction. In "Cecil Textbook of Medicine," J.B. Wyngaarden and L.H. Smith, Jr., eds., Chap. 352, 1987.
- Shope, R.E. Yellow fever. In "Cecil Textbook of Medicine," J.B. Wyngaarden and L.H. Smith, Jr., eds., Chap. 361, 1987.
- Shope, R.E. Viral hemorrhagic fevers. Introduction. In "Cecil Textbook of Medicine," J.B. Wyngaarden and L.H. Smith, Jr., eds., Chap. 360, 1987.
- Shope, R.E. Arbovirology, a look into the past and the future. Hilea Medica 8:(1) 46, 1988.
- Shope, R.E. Rabies, in Viral Infections of Humans, 3rd edition, A.S. Evans, (ed.), Chap. 19, pp. 509-523, Plenum, New York, 1989.
- Shope, R.E., Woodall, J.P. and Travassos da Rosa, A.H.P.A. The epidemiology of diseases caused by viruses in groups C and Guama (Bunyaviridae), in The Arboviruses: Epidemiology and Ecology, Vol III, T.P. Monath (ed.), CRC Press, Boca Raton, Florida, pp. 37-62, 1989.
- Schwan, T.G., Oprandy, J.J., and Main, A.J. Virus infecting <u>Argas</u> ticks associated with California gulls breeding on islands in Mono Lake, California. J. Med. Entomol. 25:381-387, 1988.
- Tesh, R.B. The genus <u>Phlebovirus</u> and its vectors. Ann. Rev. Entomol. 33:169-181, 1988.
- Tesh, R.B. Arboviruses of Central Asia and the Soviet Union. In "Textbook of Pediatric Infectious Diseases, 2nd Edition, R.D. Feigin and J.D. Cherry, editors. W.B. Saunders Co., Philadelphia, 1987, pp. 1494-1502.
- Tesh, R.B. Undifferentiated fevers: dengue, phlebotomus fever, Rift Valley fever, West Nile fever and fevers caused by alphaviruses. In: Cecil Textbook of Medicine, 18th edition, J.B. Wyngaarden and L.H. Smith, editors, W.B. Saunders Co., Philadelphia, pp. 1816-1819, 1988.

Tesh, R.B., Boshell, J., Modi, G., Morales, A., Young, D., Corredor, A., Ferro, C., Rodriguez, C. de, Walters, L.L. and Gaitan, M. Natural infection of humans, animals and phlebotomine sand flies with the Alagoas serotype of vesicular stomatitis virus in Colombia. Am. J. Trop. Med. Hyg. 36:653-666. 1987.

Tesh, R.B., Boshell, J., Young, D.G., Morales, A., Ferro de Carrasquilla, C., Corredor, A., Modi, G.B., Travassos da Rosa, A.P.A., McLean, P.G., Rodriguez, C. and Gaitan, M.O. Characterization of five new phleboviruses recently isolated from sand flies in tropical America. Am. J. Trop. Med. Hyg. 40:529-533, 1989.

Tesh, R.B., Chen, W.R. and Catuccio, D. Comparative rates of digestion of albumin, IgG, IgM and complement (C3) in human blood after ingestion by mosquitoes (Acdes albopictus) and sand flies (Phlebotomus papatasi). Am. J. Trop. Med. & Hyg. 39:127-130, 1988.

Tesh, R.B. and Modi, G.B. Maintenance of Toscana virus in <u>Phlebotomus</u> perniciosus by vertical transmission. Am. J. Trop. Med. Hyg. 36:199-203, 1987.

Tesh, R.B. and Duboise, S.M. Viremia and immune response with sequential phlebovirus infections. Am. J. Trop. Med. Hyg. 36:662-668, 1987.

Tesh, R.B. Phlebotomus fevers, in The Arboviruses: Epidemiology and Ecology, Vol IV, T.P. Monath (ed.), CRC Press, Boca Raton, Florida, pp. 15-27, 1989.

Tesh, R.B. The epidemiology of phlebotomus (sandfly) fever. Israel J. Med. Sci. 25:214-217, 1989.

Zeller, H.G., Karabatsos, N., Calisher, C.H., Digoutte, J.P., Murphy, F.A., and Shope, R.E. Electron microscopy and antigenic studies of uncharacterized viruses. I. Evidence suggesting the placement of viruses in families Arenaviridae, Paramyxoviridae, or Poxviridae. Arch. Virol. 108:191-209, 1989.

Zeller, H.G., Karabatsos, N., Calisher, C.H., Digoutte, J.P., Cropp, C.B., Murphy, F.A. and Shope, R.E. Electron microscopic and antigenic studies of uncharacterized viruses. II. Evidence suggesting the placement of viruses in the family Bunyaviridae. Arch. Virol. 108:211-217, 1989.

PROFESSIONAL PERSONNEL RECEIVING GRANT SUPPORT:

Susan Brown, Ph.D. Dennis L. Knudson, D.Phil. Peter W. Mason, Ph.D. Robert E. Shope, M.D.